Effect of Seed Priming with Some Plant Leaf Extract on Seedling Growth Characteristics and Root Rot Disease in Tomato

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(Received: April 5, 2015 Reviewed: July 23, 2015; Accepted: May 2, 2016)

Abstract: Tomato is one of the important vegetable crops. The problem of seedling establishment is found in tomato due to several soil borne diseases. One of them is root rot caused by *Fusarium oxysproum*. There are many chemical methods available to control this disease, but use of chemicals deplete the soil micro-environment and causes soil and water pollution and also do not fit within the framework of 'Organic farming'. Seed priming with certain phytochemicals may be an economic and ecofriendly alternative to such chemicals. In present study we primed tomato seeds with leaf extract of six different plants (White musale, Periwinkle, Neem, Wood apple, Lantana and White cedar). Different leaf extracts of dose of 2% was taken independently for seed priming. We found that priming with White musale, Periwinkle, Neem and wood apple leaf extract had an improvement in different seed and seedling growth parameters in presence of pathogen. Priming with Lantana and white cedar leaf extract showed a reduction in some of the parameters that may be due to allelopathic nature of these plants. Seed priming with leaf extract of Wood apple exhibited maximum survival rate (76.50 %) followed by Neem (68.46 %) and White Musale (52.60 %).

Keywords: Fusarium oxysporum; plant extracts; seed priming; seedling vigour; tomato

1. Introduction

Tomato is an important vegetable crop. Production of tomato is greatly affected by different diseases. One of the many diseases is root rot in tomato causes by *Fusarium oxysproum*. It is a soil born fungus found especially in warm regions. The organism is specific for tomato and is very longlived in soil (Suárez-Estrellaet *et al.*, 2004). Various methods are tried to control this disease organically. Seed priming with phytochemicals can also be a method to increase the seed vigour of seeds and can improve performance of seeds in presence of pathogen.

The rationale of seed priming is to lessen the time between planting and emergence and to protect seeds from biotic and abiotic factors during critical phase of seedling establishment and to synchronize emergence, which leads to uniform stand as well as improved yield (Krishnaswami and Srimarthi, 2001). Priming is controlled hydration technique and enhances seed performance by increasing germination rate and uniformity resulting in faster and better seedling development in various crops (Powell et al., 2000, Afzal et al., 2008). Leaves of the plant contain alkaloids and phenolic compounds which protect the plants against pathogens and also produce antioxidant activity (Satish et al., 2007). In the present study we use leaf extracts from six different plants for priming and emphasized on the improvement in germination, seedling growth and survival of seedling in presence of Fusarium oxysproum.

2. Materials and Method

2.1 Plant Material

Present study was carried out using tomato seeds var. Money Maker. The fresh leaves of the White Musale (*Chlorophytum borivillianum*), Periwinkle (*Vinca rosea*), Wood apple (*Aegle marmelos*), Neem (*Azadirachta indica*), White cedar (*Melia azedarach* L) and Lantana (*Lantana camara*) were collected and dried separately under shade.

2.2 Preparation of Leaf Extracts and Dose Standardization

Leaf extract was prepared using cold extraction method. The dried leaves were

powdered using electric grinder. First we standardize the dose of medicinal plants leaf extract. For this 1%, 2% and 4% solutions from the respective leaf powder were made in single distilled water. These solutions were kept at 30°C temperature for 48 hours. After 48 hours the leaf extracts were filtered by two layer of muslin cloth to remove unwanted material and leaf debris. Seeds were soaked independently in different leaf extracts at room temperature for four hours. After soaking seeds were air dried and put on towel paper for germination test. The seeds primed with 2% and 4% solution of leaf extract showed better results than 1%, however difference between priming with 2% and 4% leaf extract were not pronounced therefore we used 2% leaf extract for further experiment.

2.3 Assessment of Various Growth Parameters

Pot experiment was conducted to assess the effect of seed priming on disease suppression. The soil was inoculated with 500mg/kg inoculums of *Fusarium oxysporum*. Three controls were taken for the experiment, (1) Positive control where the seeds were treated with fungicide and the soil was inoculated with pathogen, (2) Negative control where the seeds were not given any treatment but the soil was inoculated with pathogen and (3) Normal control where neither the soil was inoculated nor the seeds were treated.

2.4 Data Analysis

Observations on germination percentage, speed of germination, germination value, root length, shoot length, fresh weight, dry weight, mortality rate were recorded and seedling vigour I and II were calculated. The data was statistically analysed using ANO-VA to calculate the magnitude of F value.

3. Results and Discussion

In the present study seed priming with different plants extract showed significant increase in different parameters over control. Seed priming with Chlorophytum extract was found best among all the treatments for different morphological and physiological parameters Maximum speed of germination was observed with the seed primed with Chlorophytum with 76.89% increase over control (Figure 1). Improved Germination percentage was recorded in all the treatments. Germination percentage ranged from 68 to 90% (Table 1). Neem leaves shows a moderate effect on the germination and seedling growth in comparison to other treatments

This increase in speed of germination and germination percentage may be due to the modification of physiological and biochemical nature of seed embryo and its associated structures, i.e. pre –enlargement of the embryo (Austin *et al.*, 1969) and biochemical changes like enzyme activation, Gibberellins like substances (Lee and Kim, 2000, Basra *et al.*, 2005) may release during the II phase of germination which triggers the synthesis of hydrolytic enzymes that causes the early availability of high energy compounds and vital biomolecules to the germinating seedling (Renugadevi and Vijayageetha, 2006).

Maximum seedling length, Dry weight, seedling vigour I and seedling vigour II were found for treatment T1 followed by T3 (Table 1). The increase in dry weight was claimed to be due to enhanced lipid utilization and enzyme activity due to the presence of bioactive substances in the leaf extracts (Rathinavel and Dharmalingam, 1999) and development of seedling to reach autotropic stage and enabling them to produce relatively more quantity of dry matter which discerning the cause for the hike in vigour index by hardening treatment. It is generally assumed that the active constituents which are contributing to these anti-fungal properties

Treatments	Speed of germination (plant /Day)	Germination percentage	Seedling length (cm)	Fresh weight (gm)	Dry weight (gm)	Seedling vigour I	Seedling vigour II	Mortality (%)
White Musale	8.65	90.83	10.30	0.57	0.06	481.75	5.62	47.33
Periwinkle	7.52	84.17	9.97	0.46	0.06	418.88	4.64	56.33
Wood apple	6.03	88.33	10.26	0.58	0.06	464.61	5.59	23.33
Neem	6.32	88.33	9.96	0.54	0.06	437.56	5.11	32.67
White cedar	4.68	89.17	9.34	0.59	0.05	476.35	4.16	58.00
Lantana	4.80	76.67	10.73	0.61	0.05	439.25	3.58	63.67
Control	5.54	71.67	8.75	0.44	0.04	340.75	2.74	35.55
Positive control	8.33	93.33	10.33	0.54	0.05	497.92	4.98	12.67
Negative control	4.89	68.33	7.16	0.40	0.03	146.65	2.16	70.67
cd at 5%	1.0102	4.74	0.80	0.12	0.01	71.88	1.02	5.46

 Table 1. Effect of seed priming on various seedling growth parameters and disease suppression in pot experiment.

of the extracts are phyto-chemicals. Physiologically active substances might have activated the embryo and other associated structures which resulted in the absorption of more water due to cell wall elasticity and development of stronger and efficient root system and that would have ultimately resulted in higher vigour index (Rangaswamy *et al.*, 1993). Many researchers also reported the benefits of seed hardening with different medicinal plants leaf extract to overcome the adverse condition (Renugadevi *et al.*, 2008; Kamaraj and Padmavathi, 2012).

Decline in speed of germination observed when the seeds were primed with *Melia* and *Lantana* (Figure 1). Ahmed *et al.* (2007) found in their experiment that different concentrations of aqueous leaf extracts of lantana caused significant inhibitory effect on germination, root and shoot elongation and development of lateral roots of receptor crops. The inhibitory effect was much pronounced in root and lateral root development rather than shoot and germination. Same results were reported by Phuwiwat *et al.* (2012) when studying the allelopathic ef-

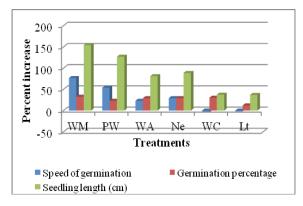


Figure 1: Increase in the different seedling growth parameters over negative control. WM- White Musale, PW-Periwinkle, WA- Wood apple, Ne- Neem, WC- White cedar and Lt- Lantana. Decrease in speed of germination observed in the seeds primed with white cedar and lantana.

fect of *Melia* leaves on the germination and seedling growth of barnyard grass. They found that both, water uptake and α -amylase activity of *E. crus-galli* were inhibited. It was concluded that leaves of *M. azedarach* contained water soluble allelochemicals and caused inhibition of both water uptake and α -amylase activity of *E. crus-galli* during germination process.

The maximum decrease in mortality rate was observed in the seeds primed with wood apple (66.99%) followed by neem (53.77%) (Table 1, Figure 2). Chlorophytum leaves are high in saponin, carbohydrate, proteins and various alkaloids (Chakraborty et al., 2014). Leaves of Vinca rosea are also high in alkaloids and phenolic compounds (Tiong et al., 2013). Neem leaves contains flavonoids, steroids, carbohydrates, glycosides, antiquinone, terpenoides and alkaloids (Raphael, 2012). Alkaloids, phnenolic and saponins compounds protect the plants against pathogens and also produce antioxidant activity. Sharma and Trivedi (2002) reported growth inhibition (72.0%) of Fusarium oxysporum f. sp cumini with the extract

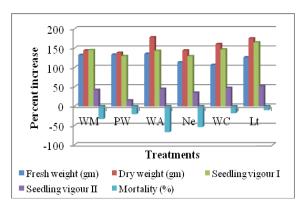


Figure 2. Increase in different planting value parameters over negative control. WM- White Musale, PW-Periwinkle, WA- Wood apple, Ne- Neem, WC- White cedar and Lt- Lantana. Minimum mortality rate was observed when the seeds were primed with leaf extract of wood apple.

of *D. stramonium*. Anti-oxidant properties scavenges the free radical activity thereby improves the germination. Saponins are readily soluble in water which may enhance the nutrient absorption (Satish *et al.*, 2007).

4. Conclusion

We can conclude from the above findings that seed priming with 2% leaf extract of Chlorophytum, Agel, Azadirachta and Vinca were found suitable to enhance seedling vigour and reduce mortality rate in presence of Fusarium oxysporum in tomato. Lantana and Melia shows a reduction in some of the physiological characters but even they could improve the seed vigour in tomato. Seed priming with these plant leaf extract is cost effective and easy to apply at farmer's field and is suitable under 'Organic farming' framework. More research is needed to understand the physiological and biochemical changes occur due to seed priming from the leaf extract of medicinal plants.

References

- Afzal, I., S.M.A. Basra and N. Ahmad. (2011). Hormonal priming induces salt tolerance in wheat through enhanced antioxidant defence system. Cereal Research Communication, 39: 334-337.
- Ahmed, R., M.B. Uddin, M.A.S.A. Khan and S.A. Mukul. (2007). Allelopathic effects of Lantana camara on germination and growth behavior of some agricultural crops in Bangladeshi. Journal of Forest Research, 18: 301-304.
- Austin, R., P.C. Longden and J. Hutchinso.

(1969). Some effects of hardening on carrot seed. Annals of Botany, 33: 883-895.

- Basra, S.M.A., M. Farooq and R. Tabassum.
 (2005). Physiological and biochemical aspects of seed vigor enhancement treatment in fine rice (*Oryza sativa* L.).
 Seed Science and Technology, 33: 623 -628.
- Chakraborthy, G.S., V. Aeri, P. Verma and S. Singh. (2014). Phytochemical and antimicrobial studies of *Chlorophytum borivilianum*. Pharmacophore, 5: 258-261.
- Kamaraj, A. and S. Padmavathi. (2012). Effect of seed ehancement treatment using leaf extract on physiological and morphological characteristics of green gram (*Vigna radiate* 1.) seed adt 3. International Journal of Current Research, 4: 110-114.
- Krishnasamy, V. and P. Srimathi. (2001).
 Seed Management of rainfed agriculture. In: Land and use planning and watershed management in Rainfed Agriculture (ed.) Balusamy, M., C.R. Chinnamuthu and A.Velayutham. Centre of Advanced studies, Department of Agronomy, Tamilnadu Agricultural University, Coimbatore. Pp. 140.
- Lee, S.S. and J.H. Kim. (2000). Total sugars, α-amylase activity and germination after priming of normal and aged rice seeds. Korean Journal of Crop Science, 45: 108 -111.
- Phuwiwat, W., W. Watcharin, L. Chamroon and T. Montinee. (2012). Inhibitory effects of *Melia azedarach* 1. leaf extracts on seed germination and

seedling growth of two weed species. Pakistan Journal of Weed Science Research, 18: 485-492.

- Powell, A.A., L.J. Yule, H.C. Jingh, and S.P.C. Groot. (2000). The influence of aerated hydration seed treatment on seed longevity as assessed by the viability equation. Journal of Experimental Botany, 51: 2031-2043.
- Rangaswamy, A., S. Purushothaman and P. Devasenapaty. (1993). Seed hardening in relation to seedling quality characters of the crops. Madras Agriculture Journal, 80: 535-537.
- Raphael, E. (2012). Phytochemical constituents of some leaves extract of Aloe vera and *Azadirachta indica* plant species. Global Advanced Research Journal of Environmental Science and Toxicology, 1: 014-017.
- Rathinavel, K. and C. Dharmalingam. (1999). Seed hardening to augument the productivity of cotton cv. LRA 5116 (*Gossypium hirsutum* L.). Madras Agriculture Journal, 86: 68 -72.
- Renugadevi, J. and V. Vijayageetha. (2006).
 Organic seed fortification in cluster bean (*Cyamopsis tetragonoloba* L.)
 TAUB. International conference on indigeneous vegetables and Legumes.
 Prospects for fighting Poverty, Hunger

and Malnutrition.

- Renugadevi, J., N. Natarajan, and P. Srimathi. (2008). Efficacy of Botanicals in improving seeds and seedling quality characteristics of Cluster bean. Legume Research, 31: 164 -168.
- Satish, S., D.C. Mohana, M.P. Ranhavendra and K.A. Raveesha. (2007). Antifungal activity of some plant extracts against important seed borne pathogens of *Aspergillus* sp. Journal of Agriculture Technology, 3: 109-119.
- Sharma, N. and P.C. Trivedi. (2002). Screening of leaf extracts of some plants for their nematicidal and fungicidal properties against *Meloidogyne incognita* and *Fusarium oxysporum*. Asian Journal of Experimental Science, 16: 21-28.
- Suárez-Estrella, F., M.C. Vargas-García, M.J. López and J. Moreno. (2004). Survival of *Fusarium oxysporum* f.sp. *melonis* on plant waste. Crop Protection, 23: 127-133.
- Tiong, S.H., C.Y. Looi, H. Hazni, A. Arya,
 M. Paydar, W. F. Wong, S. C. Cheah,
 M. R. Mustafa, and K. Awang. (2013).
 Antidiabetic and Antioxidant Properties of Alkaloids from *Catharanthus roseus* (L.) G. Don. Molecule, 18: 9770-9784.
